

VALIDATION OF THE BD PROBETEC[™] Q[×] AMPLIFIED DNA ASSAY FOR THE DETECTION OF CHLAMYDIA AND GONORRHEA IN A PUBLIC HEALTH LABORATORY SETTING

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BACKGROUND

Chlamydia trachomatis (CT) and Neisseria gonorrhoeae (GC) infections are the two most commonly reported sexually transmitted diseases.
Rapid diagnoses of CT and GC infections have significant impacts on preventing complications and reducing transmission.

INTRODUCTION

• We previously validated the BD Viper System (BD Diagnostics, Sparks, MD) for its performance of the BD ProbeTec[™] ET CT and GC Amplified DNA Assays.

- The BD Viper System is a fully automated instrument for the detection of CT and GC in urogenital specimens using the real-time Strand Displacement Amplification technology.
- The system provides for walk-away processing of the extraction, amplification, and detection of target nucleic acids, with a capacity of 184 assay results per run [(92 patient samples extracted /run) x (2 assay results/run) = 184 total results per run].

• The BD Viper System in our laboratory was recently upgraded to perform the BD ProbeTec[™] CT/GC Q[×] Amplified DNA Assay (extracted mode).

• Here, we evaluated the clinical agreement of Qx Assay when compared to the ET Assay (non-extracted mode) and their operator advantages.

MATERIALS & METHODS

• Urine specimens previously tested for CT and GC DNA by the BD ProbeTec[™] ET CT and GC Amplified DNA, as well as spiked swab samples, were tested by the BD ProbeTec[™] CT/GC Q[×] Amplified DNA Assays using the BD Viper System.

• The Q^x Assay requires the use of new Female Endocervical Specimen Collection Kit and new Male Urethral Specimen Collection Kit.

• Swab diluent spiked with AmpliTrol CT/GC (Bio-Rad Laboratories, Hercules, CA).

RESULTS

Table 1. Percentage Agreement of 236 Total Samples Tested (47 CT positive,39 GC positive)

Assay Type	Specimen Type	% Agreeement
СТ	Urine (N=186)	100%
	Swab (N=50)	100%
GC	Urine (N=186)	100%
	Swab (N=50)	100%

Sensitivity: 100%; Specificity: 100%

Table 2. Clinical Agreement of 50 Mock Swab Samples

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		Intended results/ Spiked or not	
Detection of CT		Positive	Negative
Q ^x Assay (Extracted)	Positive	22	0
	Negative	0	28
Sensitivity = 100%; Specificity = 100%			
		Intended results/ Spiked or not	
Detection of GC		Positive	Negative
Q ^x Assay (Extracted)	Positive	22	0
	Negative	0	28
Sensitivity - 100%: Specificity -100%			

Sensitivity = 100%; Specificity =100%

Table 3. Clinical Agreement of 186 Urine Samples

		ET Assay (Non-Extracted)	
Detection of CT		Positive	Negative
Q ^x Assay (Extracted)	Positive	25	0
	Negative	0	161

Sensitivity = 100%; Specificity = 100%

		ET Assay (Non-Extracted)	
Detection of GC		Positive	Negative
Q ^x Assay (Extracted)	Positive	17	0
	Negative	0	169
Sensitivity = 100%; Specificity = 100%			

Table 4. Work-flow Analysis (per 48 urine samples) of Q^{x} Assay in Comparison to the ET Assay

Q ^x Assay (Extracted)	ET Assay (Non-Extracted)
Pre-warm: 15min Cool: 15min nstrument set-up: 20min nstrument clean-up: 20min	Pre-warm: 10min Cool: 15min Centrifuge: 30min Decant, Add diluent, and Vortex: 45min Lysing: 30min Cool: 15min Instrument set-up: 15min Instrument clean-up: 15min
Fotal: 70min	Total: 175min

SUMMARY

Least amount of hands-on time: saves 105min of

technologist's time that can be re-assigned to other tasks. • Increased sensitivity and specificity according to the package insert data.

• Less interference due to interfering substances (eg. leukocytes in swab and urine samples; deodorant sprays, Bilirubin, and seminal fluid in urine samples) according to the package insert.

• Higher tolerance for blood in swab samples.

• Additional specimen source (vaginal) is FDA approved.

• No repeat testing of positives: this may reduce the cost since we repeated about 8% of the total samples tested using the non-extracted method.

CONCLUSIONS

We concluded that the performance of the BD ProbeTec[™] Q[×] Assay was equivalent to the performance of the BD ProbeTec[™] ET Assay. In addition, the Q[×] Assay offer a major advantage in terms of least amount of hands-ontime required to prepare and process specimens by the laboratory scientists. Further investigation is needed to evaluate the work-flow efficiencies and cost-effectiveness of combining the Q[×] Assay using the automated BD Viper System with lean management.

REFERENCES

• BD ProbeTec[™] Chlamydia trachomatis (CT) and Neisseria gonorrhoeae (GC) Q[×] Amplified DNA Assay Package Insert (BD 8081408 and 8081409).

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DISCLAIMER

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